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Review Article

Mesenchymal Stem Cells-Conditioned Medium (SECRETOME) in Skin Aging: A Systematic Review

WINAWATI EKA PUTRI^{1,2,3}, ANANG ENDARYANTO², FEDIK ABDUL RANTAM^{4,5}, CITA ROSITA SIGIT PRAKOESWA²

¹Post Graduate Doctoral Program, Faculty of Medicine, Universitas Airlangga, Indonesia.

²Airlangga University Faculty of Medicine, Dermatology and Venereology, Surabaya, Indonesia.

³University of Nahdlatul Ulama Surabaya, Dermatology and Venereology, Surabaya, Indonesia.

⁴Institute of Tropical Disease, Airlangga University, Stem Cell Laboratory, Surabaya, Indonesia.

⁵Department of Microbiology, Faculty of Veterinary Medicine, Airlangga University, Virology and Immunology Laboratory, Surabaya, Indonesia.

*Corresponding Author

Email ID: cita-rosita@fk.unair.ac.id

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ABSTRACT

Background: The skin aging is the most important factor that constitutes the general "well-being" and perception of "health" in humans. The mesenchymal stem cells (MSCs) have a therapeutic effect on anti-apoptosis, angiogenic, immunomodulatory and chemo-existing activity, especially in its treatment. However, the use of mesenchymal stem cell-conditioned medium (MSC-CM) as skin aging therapy requires further investigation.

Methods: The systematic literature search have the following keywords; conditioned media or conditioned medium or secretome as well as photoaging or skin aging. A total of 283 articles were reviewed from Pubmed, Wiley Online Library, Science Direct and The Cochrane Library, and only 11 articles were relevant for this systematic review.

Results: The results show that Secretome is promising in improving the skin aging process. However, due to the high heterogeneity, a meta-analysis cannot be performed.

Conclusion: The use of secretome was promising in improving skin aging process, as shown by the available preclinical studies. However, further clinical studies are needed to confirm the benefits and effects of secretome.

Keywords: mesenchymal stem cell, secretome, conditioned medium, skin aging.

INTRODUCTION

Skin aging is a complex biological process that is influenced by a combination of intrinsic and extrinsic factors¹. Furthermore, it is associated with decreased skin function, increased skin fragility, and impaired skin wound healing. In contrast to thin, atrophic, finely wrinkled and dry intrinsically aged skin, premature photoaged skin typically shows thickened epidermis, mottled discoloration, deep wrinkles, laxity, dullness and roughness¹⁻⁵. The increase in the aging of the population and the changes in skin aging that affect self-esteem leads many anti-aging therapies to be developed^{6,7}.

Stem cells (SC) therapy, which contains different types of growth factors, is commonly used for skin aging. Furthermore, they are unique populations of cells that are undifferentiated, able to renew themselves, and differentiate into different cell lineage. The mesenchymal SC (MSC) is a multipotent stromal cell that can differentiate into

various types of cells such as adipocytes, osteoblasts, chondrocytes, fibroblasts, and myoblasts. There are many types of MSC, which includes bone marrow stem cell (BMSC), umbilical cord (UC) -MSC, placenta (P) -MSC, umbilical cord blood (UCB) -MSC, amniotic fluid (AF) -MSC, and adipose stem cell (ASC)⁸. In addition, Mesenchymal stem cell produces various secreted factors like cytokines, growth factors and chemokines, which may trigger intracellular mechanism. The main functions of MSC are tissue replacement through multipotent differentiation, immunomodulatory and anti-inflammatory effects, and molecular secretion, which helps tissue repair^{9,10}. This SC is a good choice for regenerative medicine because it is easily obtained (bone marrow, adipose and UCB) and has low immunogenicity⁹.

In recent years, conditioned medium (CM) or SC-derived secretome has received more attention in the field of regenerative medicine and has generally

shown good results. The use of secretome has several advantages when compared to SC, as CM can be manufactured, freeze-dried, packaged, and transported more easily. Moreover, as it is devoid of cells, there is no need to match the donor and the recipient to avoid rejection problems¹¹. The secretome contains many growth factor, cytokines, chemokines, angiogenic factors, microvesicle and exosome. Therefore, SC-derived CM have a promising prospect to be produced as pharmaceuticals for regenerative medicine^{11,12}. In addition, its potential role as a cosmetic ingredient is being explored¹³.

A previous *in vitro* study showed that medium conditioned with adipose stem cells (ASC-CM) can increase the expression of collagen type I mRNA, type III collagen and elastin in senescence human dermal fibroblast (HDF). However, this ability were reduced, as the passage of HDF increased¹⁴. Another study conducted by Li et al showed anti photoaging activities of this ASC-CM on human keratinocytes and fibroblasts exposed to UVB light. Inhibition of phosphorylation of the MAPK/ AP-1 pathway, decrease in NF- κ B activation and downregulation of HO-1 activation as well as increase in expression of TGF- β and Smad 2/3 proteins were reported¹⁵. Therefore, the purpose of this study literature is to know the effects and

benefits of MSC-CM or secretome for skin aging therapy.

METHODS

Eligibility criteria

The inclusion criteria used an experimental study in English with human and animal study groups with skin aging (intrinsic and extrinsic factors), given the intervention in the form of the application of MSC-CM/secretome. The expected outcome was the clinical and histological outcome as the primary outcome. Non-English studies, duplicates, review studies, and irrelevant articles as exclusion criteria.

Literature search and study selection

The integrated search process was carried out in accordance with the instructions of the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines. The article was identified from searches of PubMed (MEDLINE), the Cochrane Library, Science Direct, and Wiley Online Library on June 9th, 2020. The terms used as the search keywords contains at least two words: "conditioned media" or "conditioned medium" or secretome and skin aging or photoaging. The authors (W.E.P and C.R.S.P) then reviewed the feasibility of the article on the basis of inclusion and exclusion criteria, the due diligence was conducted by the author in a discussion and independent manner.

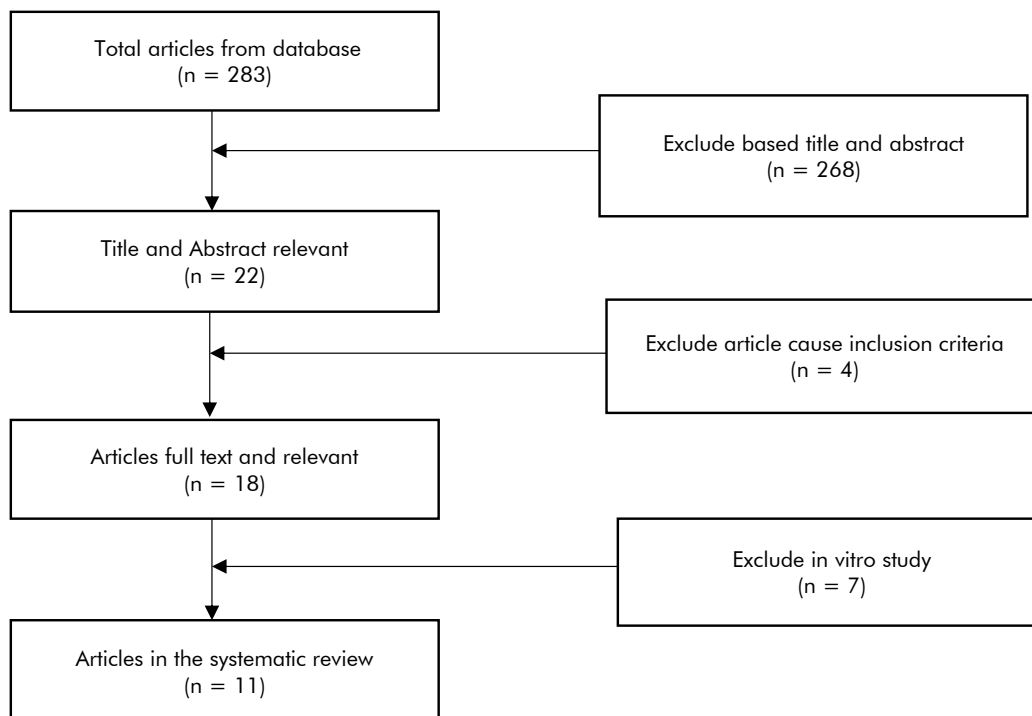


Fig.1: Prisma Flowchart

Methodological quality assessment and risk of bias

Two authors (W.E.P and C.R.S.P) in determining the assessment of methodological quality and risk of bias by combining animal testing guidelines: Reporting on in vivo Experiments (ARRIVE) guidelines with Consolidating Reporting of Trials¹⁶.

Data extraction and synthesis

The data extraction was performed by two authors (W.E.P and C.R.S.P) that were independent of each study and were in agreement with all included studies. Then, the following data were extracted: study design, human model and type of animal model for in vivo studies; establishment process for

animals or human in the included studies; type and specific donor of MSCs; the isolation process for the CM; interventions; comparison; duration of follow-up; main outcome for the in vivo studies and the results; any significant differences from control or baseline; and other outcomes. For in vivo studies, any adverse reactions were reviewed, clinical outcomes, and histological outcomes. The authors agreed to classify types of MSCs into BMSC, ASC, UCB-MSC, AF-MSC and AMSC. The data collection of in vivo study and clinical study outcomes are shown in Table 1. A meta-analysis could not be carried out due to high heterogeneity data.

Table 1: The studies overview

Author	Type of MSC	Level CM	Population	Type of Study	Sample	Intervention	Controlled
Sohn et al ¹⁷	EPC	CM	Human	Clinical study In vitro	25 Korean women (29-69 years old) with mild to moderate wrinkles. MSCs were differentiated to EPCs for 10–21 days. Conditioned medium was also collected from undifferentiated MSC.	5% EPC-CM was applied twice a day on both sides of the face for 4 weeks. NHDF cells were pretreated with each CM for 24 h, followed by exposure to 600 μ m H ₂ O ₂ for 24 h.	N/A NHDF cells were exposed to 600 μ m H ₂ O ₂ for 24 h.
Prakoewa et al ¹⁸	AMSC	CM	Human	Clinical study	48 subjects aged 41-60 years old and receive priming with 0,025% tretinoin cream for 2 weeks.	subjects were treated with microneedling plus 3 mL of AMSC-CM for 3 times with an interval of 2 weeks.	4 subjects were treated with microneedling plus 3 mL of NS; for 3 times with an interval of 2 weeks.

Author	Type of MSC	Level CM	Population	Type of Study	Sample	Intervention	Controlled
Kim et al ¹⁹	AD, BM, and UCB	CM	Human	Clinical study In vitro	22 volunteers (18–55 years-old women) Conditioned media (CM) of MSCs and HDFs were collected. The conditioned media were measured with GDF-11 ELISA kit.	USC-CM in cream base were treated daily to their skin. HDFs (1×10 ³ cells/well) were seeded in 96-well plates and cultured for 24 h in KSB-3 medium. After washing, the medium was replaced by HDF-CM, AD-MSC-CM and UCB-CM.	N/A KSB-2 media.

Author	Type of MSC	Level CM	Population	Type of Study	Sample	Intervention	Controlled
				In vitro	It was irradiated with UVB source, with dosage 10 mJ/cm ² , once a day for 5 days.	and DA-CM-treated SIPS-HDFs (HDFs cultured in DA-CM and exposed to UVB irradiation).	HDFs (HDFs cultured in serum-free DMEM), SIPS-HDFs (HDFs cultured in serum-free DMEM and exposed to UVB irradiation).
El-Donyati et al ²²	AF-MS-C	CM	Human	Clinical Study	10 volunteers (3 males and 4 females) age 41-60 years old with facial aging	5 sessions of skin needling, 2 weeks apart, AF-MS-C-CM was added topically to the right side only.	Left side, only skin needling.
Lee et al ²³	ESC-EPC	CM	Human	Clinical study	25 participants were 41-64 years old and had Fitzpatrick Skin Type III or IV.	5 treatment sessions of microneedle with hESC-EPC CM were repeated at 2-week intervals.	5 treatment sessions of microneedle with saline were repeated at 2-week intervals.
Amirthalingam et al ¹³	BMSC	CM	Mice	In vivo	6 to 8 weeks old nude mice (NU/ NU)	All mice were irradiated with UVB at a dose of 150 mJ/cm ² , once daily, for seven days and given CM.	Not irradiated with UVB
				In vitro	HFF cells were photo-damaged by UVB irradiation (300 mJ/cm ²).	HFF treated with 0.25%, 0.5% and 1% for 48 h.	
Kim et al ¹⁹	ASC	CM	Human	Clinical Study	Female adults aged 30-60 years were selected.	Cream containing 3D cultured ADMSCs-CM was applied on: Left lower forearm Left upper forearm after treated with SLS Face HDF was treated with 2D or 3D cultured ASC-CM	
				In vitro	HDF were obtained from males of 20 years old or under		Negative control was cultured with basal medium without serum. Positive control was cultured in a basal

Author	Type of MSC	Level CM	Population	Type of Study	Sample	Intervention	Controlled
							medium supplemented with 0.04%.
Wang et al ²⁴	ASC	Protein extract of CM	Human	Clinical Study	30 females between 40 and 63 years (skin type III and IV),	Protein extracts from ADSC-CM were applied via microneedles into the skin with interval 2 weeks (W0, W2, W4, W6, W8, W10).	Ultrapure water was applied via microneedles in control group.
Kwon et al ²⁵	BMSC	CM	Mice	In vivo In vitro	48 photoaged female SKH-1 hairless mice, 6 weeks old Human dermal fibroblasts	Topical BMSC-CM was applied on dorsal skin mice 3x/week for 8 weeks. Treated by MSC-CM (0,1%, 1% and 10%) for 24 and 48 h	Vehicle solution (polyethylene glycol: ethanol 7:3) was applied on dorsal skin mice 3x/week for 8 weeks NA

Table 2: In vivo studies and clinical studies results

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
Sohn et al ¹⁷	4 weeks	Crow's feet. The elevation of skin surface on the right cheek. The texture small (Ra) values of the right cheek.	Improved significantly during clinical test period.	Yes (p<0.001)	N/A	N/A	N/A
Prakoeswa et al ¹⁸	4 weeks and 8 weeks	Pore (baseline) Pore 4 weeks Pore 8 weeks Wrinkle (baseline) Wrinkle 4 weeks Wrinkle 8 weeks Spot (polarized) (baseline) Spot (polarized) 4 weeks Spot (polarized) 8 weeks Spot (UV) baseline	NS: 49.63 ± 11.193 AMSC-CM: 53.17 ± 4.565 NS: 49.58 ± 6.903 AMSC-CM: 51.42 ± 4.745 NS: 50.29 ± 4.467 AMSC-CM: 48.46 ± 6.171 NS: 12.13 ± 7.011 AMSC-CM: 13.92 ± 6.639 NS: 12.29 ± 6.196 AMSC-CM: 12.42 ± 6.413 NS: 11.67 ± 5.498 AMSC-CM: 9.29 ± 5.279 NS: 32.79 ± 8.968 AMSC-CM: 34.96 ± 6.557 NS: 34.46 ± 7.852 AMSC-CM: 32.71 ± 7.474 NS: 34.08 ± 9.016 AMSC-CM: 30.46 ± 8.335 NS: 14.54 ± 8.748	Yes Yes Yes Yes	N/A N/A N/A N/A N/A	Side effect	Erythema, urticaria (resolved after 2 days with topical 1% hydrocortisone cream).

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
		Spot (UV) 4 weeks	AMSC-CM: 17.17 ± 8.646 NS: 13.50 ± 5.934 AMSC-CM: 13.67 ± 5.880		N/A		
		Spot (UV) 8 weeks	NS: 13.04 ± 6.196 AMSC-CM: 12.46 ± 7.581		N/A		
		Skin tone (baseline)	NS: 38.58 ± 8.717 AMSC-CM: 39.54 ± 7.028	No	N/A		
		Skin tone 4 weeks	NS: 38.58 ± 8.677 AMSC-CM: 38.96 ± 7.624		N/A		
		Skin tone 8 weeks	NS: 36.67 ± 7.750 AMSC-CM: 36.92 ± 6.788		N/A		
Kim et al ¹⁹	2 weeks and 4 weeks	Dermal density baseline 2 weeks 4 weeks	14.20 ± 0.09 14.33 ± 0.08 14.55 ± 0.10	- No Yes	N/A	Adverse reaction	No adverse reaction
		Skin wrinkle Ra baseline Ra 2 weeks Ra 4 weeks	13.62 ± 0.56 13.40 ± 0.50 13.08 ± 0.45	- No No	N/A		
		Rmax baseline Rmax 2 weeks Rmax 4 weeks	94.93 ± 4.31 92.97 ± 3.34 90.35 ± 3.44	- No Yes	N/A		
		Rz baseline Rz 2 weeks Rz 4 weeks	68.87 ± 2.49 67.10 ± 2.08 66.04 ± 2.15	- No No	N/A		
		Rp baseline Rp 2 weeks Rp 4 weeks	42.66 ± 1.19 41.70 ± 1.20 40.35 ± 1.02	- No Yes	N/A		
		Rv baseline Rv 2 weeks Rv 4 weeks	57.23 ± 3.43 56.00 ± 2.54 54.59 ± 2.70	- No No	N/A		

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
Yan Xu et al ²⁰	15 days	Erythema index Melanin index Glossymeter index TEWAmeter index Corneometer index	N/A N/A N/A N/A N/A	N/A N/A N/A N/A N/A	Control group > ASC-CM (p<0.01) Control group > PSC-CM (p>0.05) Control group > ASC-CM (p<0.01) Control group > PSC-CM (p<0.01) ASC-CM < PSC-CM (P<0.01) Control group < ASC-CM (p<0.01) Control group < PSC-CM (p<0.01) Control group > ASC-CM (p<0.05) Control group > PSC-CM (p<0.01) Control group < ASC-CM (p<0.01) Control group < PSC-CM (p<0.01)	N/A	N/A
Yang Xu et al ²¹	4 weeks and 8 weeks	Collagen amount 4 weeks Collagen amount 8 weeks Collagen Type I 4 weeks Collagen Type I 8 weeks Collagen Type III	control: 19.78% ± 2.01% DA-CM: 34.31% ± 3.44% control: 19.35% ± 2.53% DA-CM: 49.33% ± 4.78% control: 19.01% ± 2.22% DA-CM: 23.14% ± 2.43% control: 22.02% ± 3.41% DA-CM: 45.37% ± 3.07% control: 23.31% ± 1.34% DA-CM: 45.93% ± 3.50%	Yes (p<0,05) Yes (p< 0.05) Yes (p <0.05) Yes (p< 0.05) Yes (p< 0.05)	N/A N/A N/A N/A	Immunohistochemistry	MMP-1 MMP-3

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
		4 weeks Collagen Type III 8 weeks MMP-1 MMP-3	control: 23.02% ± 2.42% DA-CM: 56.94% ± 6.63% N/A N/A	Yes (p< 0.05) N/A N/A	N/A N/A N/A		
El-Domyati et al ²²	One month	Clinical Improvement Left side Right side Score Left side Very good Good Moderate Mild Right side Very good Good Moderate Mild Histological evaluation Collagen fibers Before After Elastic fiber Before After	33.3 ± 8.95 60.6 ± 9.77 Disorganized collagen bundles with increased intercellular spaces Increased and more organized collagen bundles with decreased interfibrillary spaces Dense elastotic material Elastotic material decreased, fine and well-arranged new elastic fibers	Yes (p=0.026). Yes (p= 0.019)	Yes (p<0.001) Yes (p=0.003)	Side effects	Erythema Edema Pigmentary change Ecchymosis Crusting

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
		Mean epidermal thickness Left side Before After	47.55 ± 5.52 62.18 ± 5.69	Yes (p<0.001)	No (p=0.78)		
		Right side Before After	48.19 ± 4.36 64.08 ± 4.30	Yes (p<0.001)	No (p=0.41)		
Lee et al ²³	12 weeks	Clinical assessment: Overall satisfaction scores	Control: 2.72 ± 1.45 hESC-EPC: 3.25 ± 1.26	N/A	Yes (p<0.05)	Adverse event	Mild pain and temporary erythema, mild desquamation
		Assessment for pigmentation	Control: 1.32 ± 0.62 hESC-EPC: 1.54 ± 0.57	N/A	Yes (p<0.05)		
		Assessment for wrinkle	Control: 1.49 ± 0.48 hESC-EPC: 1.92 ± 0.42	N/A	Yes (p<0.05)		
		Non-invasive measurement Pigmentation Baseline	Control: 143 ± 11.1 hESC-EPC: 138 ± 14.2		Yes (p<0.05)		
		Two weeks	Control: 136 ± 12.8 hESC-EPC: 113 ± 12.1	No (p= 0.052) Yes (p<0.05)			
		Erythema Baseline	Control: 268 ± 25.1 hESC-EPC: 271 ± 24.2		Yes (p<0.05)		
		Two weeks	Control: 243 ± 21.1 hESC-EPC: 255 ± 29.8	No (p>0,05) Yes (p<0,05)			
		Wrinkle R2 value Baseline	Control: 0.52 ± 0.07 hESC-EPC: 0.58 ± 0.1		Yes (p<0.05)		
		Two weeks	Control: 0.50 ± 0.1	No (p>0.05)			

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
		R3 value Baseline Two weeks	hESC-EPC: 0.46 ± 0.09 Control: 0.38 ± 0.06 hESC-EPC: 0.4 ± 0.1 Control: 0.34 ± 0.1 hESC-EPC: 0.31 ± 0.06	Yes ($p < 0.05$) No ($p = 0.51$) Yes ($p < 0.05$)	 Yes ($p < 0.05$)		
Amirthalingam et al ¹³	8 days	skin moisture control group UVB exposed group Formulation treated Erythema Index control group UVB exposed group Formulation treated histological Epidermal thickness Control UVB-irradiated Formulation treated Dermal thickness Control UVB-irradiated Formulation treated Epidermal hyperplasia Control UVB-irradiated Formulation treated Hyperkeratosis Control UVB-irradiated Formulation treated Infiltration of inflammatory	 49.6 ± 3.7 - 55.80 ± 1.2 54.60 ± 4.7 - 14.5 ± 1.3 47.8 ± 3.2 - 37.4 ± 2.5 N/A 286.8 ± 12.9 - 366.2 ± 14.4 265 ± 14.3 - 271.4 ± 15.9 3.06 ± 0.41 14.25 ± 1.19 10.98 ± 2.05 55.18 ± 3.64 99.21 ± 8.73 76.38 ± 4.60 - +++ + - +++ +	 N/A N/A	 Yes ($p < 0.05$) Yes ($p < 0.05$)	N/A	N/A

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
		cells in dermis Control UVB-irradiated Formulation treated	- +++ ++				
Kim et al ¹⁹	3 weeks 2 weeks and 4 weeks Right after 1 time use and after 20 minutes of artificial cool breeze	Skin irritation degree after 0.5 h 24 h 48 h Skin texture damaged by external stimuli Deep skin elasticity Ur/Ue value 2 weeks 4 weeks Wrinkle assessment 2 weeks 4 weeks Dermal density 2 weeks 4 weeks Skin moisture retention effect in artificial cold wind environment Right after 1 time use 20 minutes after cool breeze	0 0 0 Control: 3.74% Application site: 11.94% 5.97% 9.34% 5.01% 6.23% 6.97% 12.5% 543.60% 452.38%	N/A N/A Yes (p<0.05) Yes (p<0.05) Yes (p<0.05) Yes (p<0.05) Yes (p<0.05)	N/A Yes (p<0.05) N/A N/A N/A		
Wang et al ²⁴	12 weeks	Melanin index Skin color	N/A N/A	Yes (p<0.001) week 10 and week 12 No	Yes p<0.01 week 2 p<0.001 week 6, 10, 12 Yes p<0.01 week 10, 12	Facial photograph Adverse effect	N/A Erythema Dryness Desquamation Burning Itching Stinging

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
		Skin radiance	N/A	Yes p<0.05 week 2 p<0.01 week 6, 12	Yes p<0.001 week 6, 10, 12		
		Skin surface topography SEsm	N/A	Yes AAPE: p<0.01, week 6 p<0.001, week 10, 12 Control: p<0.05, week 12	No		
		SEsc	N/A	Yes AAPE: p<0.05, week 10 p<0.01, week 12 Control: p<0.05, week 10 p<0.01, week 12			
		Sew	N/A N/A	Yes AAPE: p<0.05, week 2, 6 p<0.001, week 10, 12 Control: p<0.01, week 10, 12			
		Value at each time	N/A	Yes AAPE: p<0.01, week 2 p<0.001, week 6, 10, 12 Control: p<0.05, week 10 p<0.01, week 12			
		Skin elasticity R2 value	N/A	Yes AAPE:	Yes p<0.05 week 2 p<0.01 week 6		

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
		Forehead		p<0.001, week 6, 10, 12 Control: p<0.01, week 10, 12	p<0.001 week 10, 12		
		Crow's feet		Yes AAPE: p<0.01, week 2 p<0.001, week 6, 10, 12 Control: p<0.05, week 10 p<0.01, week 12	Yes p<0.01 week 2, 12 p<0.001 week 6, 10		
		Periorbital skin relief	AAPE week 12:	Yes AAPE: p<0.05, week 2 p<0.01, week 6,10,12	Yes p<0.05 week 12		
		Ra	-3.08±2.61	Yes AAPE: p<0.05, week 2, 10 p<0.01, week 6,12	Yes p<0.01 week 12		
		Rq	-3.83±3.15	Yes AAPE: p<0.05, week 2, 6, 10 p<0.001, week 12	Yes p<0.01 week 12		
		Rz	-11.48±10.01	Yes AAPE: p<0.05, week 2, 6, 10 p<0.01, week 12	Yes p<0.05 week 12		
		Rmax	-21.66±21.90	N/A			

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
				N/A			
		Self-evaluation questionnaire		N/A	Yes p<0.01 week 2, 6, 10, 12		
		Anti aging Wrinkles	N/A				
		Wrinkles around eyes	N/A	N/A	Yes p<0.05, week 2 p<0.01 week 6, 10, 12		
		Skin more resilient	N/A	N/A	Yes p<0.05, week 2, 6 p<0.01 week 10, 12		
		Skin more elastic	N/A	N/A	Yes p<0.05, week 2, 10 p<0.01 week 6, 12		
		Lifting effect	N/A		Yes p<0.05, week 2, 6 p<0.01 week 10, 12		
		Fine wrinkles reduced	N/A	N/A	Yes p<0.05, week 2, 10,12 p<0.01 week 10, 6		
		Whitening and moisturizing					
		Feels rejuvenated	N/A	N/A	Yes p<0.05, week 10 p<0.01, week 6, 12 p<0.001, week 2		
		Skin more hydrated	N/A	N/A	Yes p<0.01, week 2, 6, 10		

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
		Skin more radiant	N/A	N/A	p<0.001, week 12 Yes p<0.05, week 2 p<0.01, week 10 p<0.001, week 6, 12		
		Skin healthier	N/A	N/A	Yes p<0.01, week 2, 6, 10 p<0.001, week 12		
		Skin more fair	N/A		Yes p<0.01, week 2, 6, 10 p<0.001, week 12		
		Overall satisfaction	N/A		Yes p<0.01, week 2, 6, 10 p<0.001, week 12		
					Yes p<0.05, week 2 p<0.01, week 6, 10 p<0.001, week 12		
Kwon et al ²⁵	8 weeks	TEWL	N/A	N/A	Yes p<0.001, normal group, adenosine and 10% MSC-CM group p<0,01, UVB, 1% MSC-CM	Immunohistochemistry Histological evaluation	Type I collagen Epidermal thickening Collagen fiber
		Skin hydration	N/A	N/A	Yes p<0.001, normal group p<0.01, 1% MSC-CM		
		Keratin content	N/A	N/A	Yes p<0.001, normal group, adenosine, 1 % MSC-CM and 10% MSC-CM group		
		Wrinkle area	N/A	N/A	Yes		

Author	Assessment of main outcome	Main outcome measure(s)	Results	Significant difference from baseline	Significant difference between groups	Other evaluation	Other outcome measure(s)
		Clinical score	N/A	N/A	<p>p<0.001, normal group, adenosine, 1 % MSC-CM and 10% MSC-CM group</p> <p>Yes</p> <p>p<0.001, normal group, adenosine, 1 % MSC-CM and 10% MSC-CM group</p>		

Assessment of methodology quality

Study characteristics: Table 1 presented an overview of the study, which describes the type of MSC and CM level used, study design, human or animal model for in vivo study, cell preparation for in vitro study, and the intervention. Table 2 mostly describes the study's outcomes.

Out of 11 studies, 8 were human-model clinical studies, and 2 studies mentioned the Fitzpatrick skin type and only 1 mentioned Glogau classification for photoaging. Others did not mention the classification. Furthermore, 3 studies in this review used mice, including nude BABL/C and, SKH-1 hairless, and nude mice (NU/NU). The aging model in animal was established by irradiating with UVB light.

The most common type of MSC used was adipose tissue (five studies), followed by bone marrow (two studies), amnion tissue, epidermal progenitor cell, human umbilical cord blood, placental tissue and amniotic fluid. Topical MSC was used in 5 studies, while injection was used in 2. Prakoeswa et al, Lee et al, Wang et al and Domyati et al used microneedle and MSC-CM compared between treatment group and the control group. Study carried out by Xu et al compared between ASC-CM or PSC-CM, which was combined with hyaluronic acid.

In vivo study outcomes: Table 2 presented In vivo and clinical study outcomes, including the main outcomes measures, the scores or results, significant difference, and also other outcome measures. Out of 11 studies, clinical outcomes were mostly reported. Sohn et al, Prakoeswa et al, Kim et al reported the significant clinical improvement from baseline, which includes Crow's feet, skin surface, skin texture; pore, wrinkle, spot, skin tone; dermal density, Rmax, Rp; and pore, wrinkle, pigment, moisture, skin elasticity, sebum U zone, and T zone, respectively¹⁷⁻¹⁹. Otherwise, Yan Xu et al only reported significance difference between the control group and the treatment group²⁰. Index of Melanin, Glossy meter, Tewameter and Corneometer of ASC-CM and PSC-CM group showed improvement than control group for 15 days. However, Erythema index, reported significance difference from baseline only in ASC-CM group. Melanin index of ASC-CM group was significantly lower than PSC-CM group. Lee et al showed that after treatment for 12 weeks with microneedle and hESC-EPC CM, clinical assessment such as overall satisfaction scores, assessment for pigmentation and wrinkle were significantly improved between control and hESC-EPC CM group. Pigmentation, wrinkle and erythema were significantly decreased in 12 weeks. The overall satisfaction reported by Wang

et al was significantly improved between group in week 2, week 6 and week 12²⁴. Xu et al, Amirthalingam et al and Kwon et al, used photoaged mice model to observe the effect of MSC-CM^{13,19,21}. Amirthalingam et al and Kwon et al, that uses both BMSC-CM, reported an improvement in clinical and histological outcomes^{13,19,21}.

Histological outcomes were reported in 4 studies, all of which showed good improvements in the histological structure of aging skin, and 2 studies showed significance difference from baseline. Xu et al reported the significant increase of collagen amount, collagen type I and collagen type III in BABL/C mice treated with DA-CM, compared with control (DMEM) group for 4 weeks and 8 weeks. Domyati et al reported that after one-month treatment, mean epidermal thickness left side (microneedle only) and right side (microneedle and AF-MSC-CM) increased significantly; however, there is no significance difference between these group.

Other evaluations performed by the authors aim to monitor side effect or adverse reaction, facial photograph and immunohistochemistry. Lee et al reported mild pain, temporary erythema and mild desquamation with microneedle. Similar to Lee et al, Prakoeswa et al and Domyati et al also used microneedle and reported erythema. Only Xu et al conducted immunohistochemistry examination. All results supported the beneficial effects of MSC-CM in the treatment of skin aging.

DISCUSSION

The medium or secretome conditioned by mesenchymal stem cells contains many growth factors, cytokines, chemokines as well as extracellular matrix proteins and exosomes. These secreted factors can play a role in tissue repair and regeneration through their paracrine effect^{10-12,26,27}. This study showed that MSC-CM is always associated with significantly better clinical and histological outcomes. In addition, 11 clinical or in vivo studies showed improvement of clinical outcomes with or without histological outcomes. Some studies performed in vitro study to support in vivo studies.

An in vitro study performed by Xu et al, Sohn et al, and Pan et al, all reported an increase in superoxide dismutase (SOD). The combined laser therapy and ASC-CM conducted by Xu et al also showed a decrease in malondialdehyde (MDA) and increased in the expression of Wnt/ β -catenin, which plays a role in collagen formation for the improvement of skin aging²¹. Sohn et al did culture of HDF with H₂O₂ and reported that hEPC-CM and MSC-CM have similar protective effects on SOD and glutathione peroxidase (GPx),

which significantly increase type I procollagen also inhibited hydrogen peroxide-induced phosphorylation of proteins in mitogen activated protein kinase (MAPK) signalling and play a role in skin aging¹⁷. Parado et al²⁸ demonstrated that AMSC-CM increased catalase (CAT) and decreased malondialdehyde (MDA) also blocked cell cycle. These could delay oxidative stress-induced premature senescence²⁸. Kim et al reported that cell migration and collagen type I and type III synthesis were increased in the UCB-MS-CM group than in HDF-CM and ASC-CM. They also reported that GDF11 (rejuvenation factor) secretion was the highest in UCB-MS-CM, and therefore improving the skin wrinkles¹⁹. GDF-11 or bone morphogenetic protein-11 (BMP-11), member of TGF- β family, decreases with age. Although not high, ASC also secretes GDF-11, which can improve proliferation, differentiation of cell and migration and secretion of ECM that is important for skin aging²⁹.

An in vitro study performed by Balasubramanian et al demonstrated that BMSC-CM showed beneficial effects on the damaged fibroblasts caused by oxidative stress or UV radiation³⁰. The secretome was able to restore fibroblast proliferation and migration, also an increase in synthesis of ECM²⁶. They also reported an increase in the cyclin B1 levels, which decreases in senescence cell. Another study supported BMSC-CM as anti-aging, conducted both in vivo and invitro by Amirthalingam et al reported that the cyclobutene pyrimidine dimers (CPD) formation, which cause primary lesion in UV-irradiated DNA, was prevented by BMSC-CM treatment. They showed that these CM extensively microscopically and macroscopically protected mice skin from toxic UVB-irradiation¹³.

Sohn et al, Prakoeswa et al, and Kim et al agreed that MSC-CM can improve the appearance of wrinkle. A study by Prakoeswa et al using AMSC-CM with microneedling showed its capability to improve clinical photoaging and it was a promising option for rejuvenation therapy¹⁸. Another in vivo study conducted by Kim et al showed that topical treatment of UCB-CM showed anti-wrinkle effect and significantly increased dermal density in photoaged women¹⁹. A study by Xu et al showed that ASC-CM and PSC-CM improve skin aging for 20 days. Indeed, every type of MSC has shown improvement in skin aging. Secretome composition varies between donors depending on the type of cultured MSC²⁶. Side effects of the application of the CM were observed only in 4 studies.

This study had several limitations. First, out of 11 studies, only 2 reported clinical and histological results with randomized subjects. Second, these

studies used different cell sources, research subject, and delivery methods, which clearly leads to different outcomes. The studies used in our review varies considerably in terms of MSC sources (epidermal progenitor cell, amniotic membrane, adipose, bone marrow, umbilical cord blood, placental, and amniotic fluid), research subjects (human, mice), and delivery methods (with or without device-microneedle, laser, with or without additional material-hyaluronic acid). Finally, the clinical outcomes and the device used in measuring the same index also varies between studies. Therefore, it was impossible to perform a quantitative analysis with the included studies. Further research is needed to decide which type of MSC-CM will be the best choice for skin aging and to know more about mechanism of MSC-CM in skin aging improvement.

CONCLUSION

The use of secretome was promising in improving skin aging process, as shown by the available preclinical studies; however, further clinical studies are needed to choose which type of MSC is superior for the improvements of skin aging.

CONFLICT OF INTEREST

There is no conflict of interest in this study literature.

REFERENCES

1. Mohiuddin AK. Skin Aging & Modern Edge Anti-Aging Strategies. *Int J Dermatology Ski Care*. 2019;19:8-62.
2. Chung J, Cho S, Kang S. Why Does the Skin Age? Intrinsic Aging, Photoaging, and Their Pathophysiology. In: Rigel D, Weiss R, Lim H, Dover J, eds. Marcel Dekker, Inc; 2004.
3. Assaf H, Adly M, Hussein M. Aging and Intrinsic Aging: Pathogenesis and Manifestations. In: Farage M, Miler K, Maibach H, eds. *Textbook of Aging Skin*. Springer; 2010.
4. Rittié L, Fisher GJ. Natural and sun-induced aging of human skin. *Cold Spring Harb Perspect Med*. 2015;5(1):1-14. doi:10.1101/cshperspect.a015370
5. Garg C, Khurana P, Garg M. Molecular mechanisms of skin photoaging and plant inhibitors. *Int J Green Pharm*. 2017;11(02):217-232.
6. De Haes P, Garmyn M, Verstuyf A, et al. 1,25-Dihydroxyvitamin D3 and analogues protect primary human keratinocytes against UVB-induced DNA damage. *J Photochem Photobiol B Biol*. Published online 2005. doi:10.1016/j.jphotobiol.2004.09.010
7. Mohiuddin AK. Skin Aging & Modern Age Anti-aging Strategies. *Glob J Med Res*. Published online 2019. doi:10.34257/gjmr/vol19is2pg15

8. Hasegawa T, Ikeda S. Mesenchymal Stem Cells for the Treatment of Skin Diseases. *AIMS Cell Tissue Eng.* 2017;1(2):104-117. doi:10.3934/celltissue.2017.2.104
9. Lavoie JR, Rosu-Myles M. Uncovering the secretome of mesenchymal stem cells. *Biochimie.* 2013;95(12):2212-2221. doi:10.1016/j.biochi.2013.06.017
10. Maumus M, Jorgensen C, Noël D. Mesenchymal stem cells in regenerative medicine applied to rheumatic diseases: Role of secretome and exosomes. *Biochimie.* Published online 2013. doi:10.1016/j.biochi.2013.04.017
11. Pawitan JA. Prospect of Stem Cell Conditioned Medium in Regenerative Medicine. *Biomed Res Int.* 2014;2014:7-9. doi:10.1155/2014/965849
12. Xia J, Minamino S, Kuwabara K, Arai S. Stem cell secretome as a new booster for regenerative medicine. *Biosci Trends.* 2019;13(4):299-307. doi:10.5582/bst.2019.01226
13. Amirthalingam M, Bhat S, Dighe PA, Seetharam RN. Human Mesenchymal Stromal Cells-Derived Conditioned Medium Based Formulation for Advanced Skin Care: in vitro and in vivo Evaluation. *J Stem Cell Res Dev Ther.* 2019;4:12.
14. Wang T, Guo S, Liu X, Xu N, Zhang S. Protective effects of adipose-derived stem cells secretome on human dermal fibroblasts from ageing damages. *Int J Clin Exp Pathol.* 2015;8(12):15739-15748.
15. Li L, Ngo HTT, Hwang E, et al. Conditioned medium from human adipose-derived mesenchymal stem cell culture prevents uvb-induced skin aging in human keratinocytes and dermal fibroblasts. *Int J Mol Sci.* 2020;21(1). doi:10.3390/ijms21010049
16. Abbas Q, Emre Celebi M, Fondón I. Computer-aided pattern classification system for dermoscopy images. *Ski Res Technol.* Published online 2012. doi:10.1111/j.1600-0846.2011.00562.x
17. Sohn SJ, Yu JM, Lee EY, et al. Anti-aging Properties of Conditioned Media of Epidermal Progenitor Cells Derived from Mesenchymal Stem Cells. *Dermatol Ther (Heidelb).* Published online 2018. doi:10.1007/s13555-018-0229-2
18. Prakoeswa CRS, Pratiwi FD, Herwanto N, et al. The effects of amniotic membrane stem cell-conditioned medium on photoaging. *J Dermatolog Treat.* 2019;30(5):478-482. doi:10.1080/09546634.2018.1530438
19. Kim YJ, Seo DH, Lee SH, et al. Conditioned media from human umbilical cord blood-derived mesenchymal stem cells stimulate rejuvenation function in human skin. *Biochem Biophys Reports.* 2018;16(October):96-102. doi:10.1016/j.bbrep.2018.10.007
20. Xu Y, Guo S, Wei C, et al. The Comparison of Adipose Stem Cell and Placental Stem Cell in Secretion Characteristics and in Facial Antiaging. *Stem Cells Int.* Published online 2016. doi:10.1155/2016/7315830
21. Xu Y, Zhang JA, Xu Y, et al. Antiphotaging Effect of Conditioned Medium of Dedifferentiated Adipocytes on Skin in Vivo and in Vitro: A Mechanistic Study. *Stem Cells Dev.* Published online 2015. doi:10.1089/scd.2014.0321
22. El-Domyati M, Moftah NH, Nasif GA, Ameen SW, Ibrahim MR, Ragaie MH. Facial rejuvenation using stem cell conditioned media combined with skin needling: A split-face comparative study. *J Cosmet Dermatol.* Published online 2020. doi:10.1111/jocd.13594
23. Lee HJ, Lee EG, Kang S, Sung JH, Chung HM, Kim DH. Efficacy of microneedling plus human stem cell conditioned medium for skin rejuvenation: A randomized, controlled, blinded split-face study. *Ann Dermatol.* Published online 2014. doi:10.5021/ad.2014.26.5.584
24. Wang X, Shu X, Huo W, Zou L, Li L. Efficacy of protein extracts from medium of Adipose-derived stem cells via microneedles on Asian skin. *J Cosmet Laser Ther.* Published online 2018. doi:10.1080/14764172.2017.1400171
25. Kwon TR, Oh CT, Choi EJ, et al. Conditioned medium from human bone marrow-derived mesenchymal stem cells promotes skin moisturization and effacement of wrinkles in UVB-irradiated SKH-I hairless mice. *Photodermatol Photoimmunol Photomed.* Published online 2016. doi:10.1111/phpp.12224
26. Rengasamy M, Singh G, Fakhruzi NA, et al. Transplantation of human bone marrow mesenchymal stromal cells reduces liver fibrosis more effectively than Wharton's jelly mesenchymal stromal cells. *Stem Cell Res Ther.* Published online 2017. doi:10.1186/s13287-017-0595-1
27. Sagaradze G, Grigorieva O, Nimiritsky P, et al. Conditioned medium from human mesenchymal stromal cells: Towards the clinical translation. *Int J Mol Sci.* Published online 2019. doi:10.3390/ijms20071656
28. Parrado C, Mercado-Saenz S, Perez-Davo A, Gilaberte Y, Gonzalez S, Juarranz A. Environmental Stressors on Skin Aging. Mechanistic Insights. *Front Pharmacol.* Published online 2019. doi:10.3389/fphar.2019.00759
29. Mazini L, Rochette L, Malka G. Adipose-Derived Stem Cells (ADSCs) and Growth Differentiation Factor 11 (GDF11): Regenerative and Antiaging Capacity for the Skin. In: *Regenerative Medicine.* ; 2020. doi:10.5772/intechopen.91233
30. Balasubramanian P, Howell PR, Anderson RM. Aging and Caloric Restriction Research: A Biological Perspective With Translational Potential. *EBioMedicine.* Published online 2017. doi:10.1016/j.ebiom.2017.06.015